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THE 2016 GLUTEN-FREE STAKEHOLDER UPDATE & PLANNING SESSION PRESENTED IN COOPERATION WITH:



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# CAPABILITIES AND CHALLENGES FOR DETECTION OF GLUTEN BY IMMUNOASSAY

Scott Radcliffe, Technical Support Scientist, Romer Labs  
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# ALLERGEN DETECTION METHODS

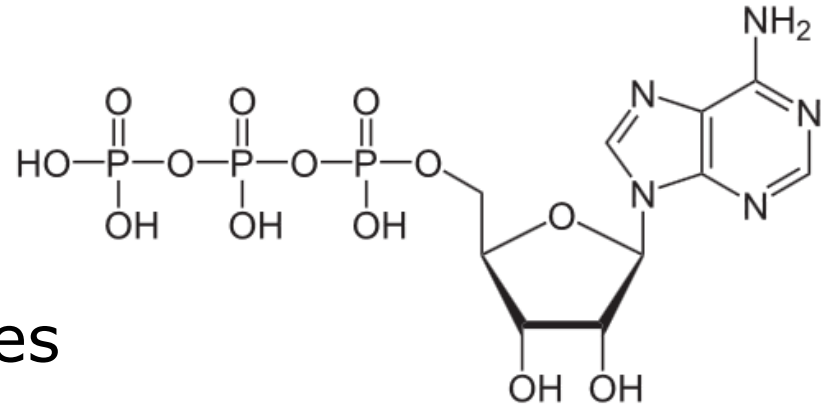


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# ATP SWAB

- Energy transport molecule found in all living cells
- Rapid & inexpensive, requires luminometer
- Result correlates surface cleanliness
- ATP is not a protein or allergen
- Negative ATP test  $\neq$  Negative for allergen protein

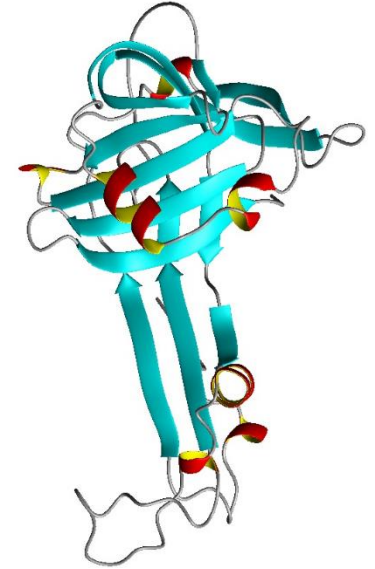


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# PROTEIN SWAB

- Rapid, inexpensive
- Chemical rxn with protein residue causes color change from green to purple
- Since allergens are protein, better indicator allergens removed than ATP
- Non-specific, less sensitive than allergen immunoassays

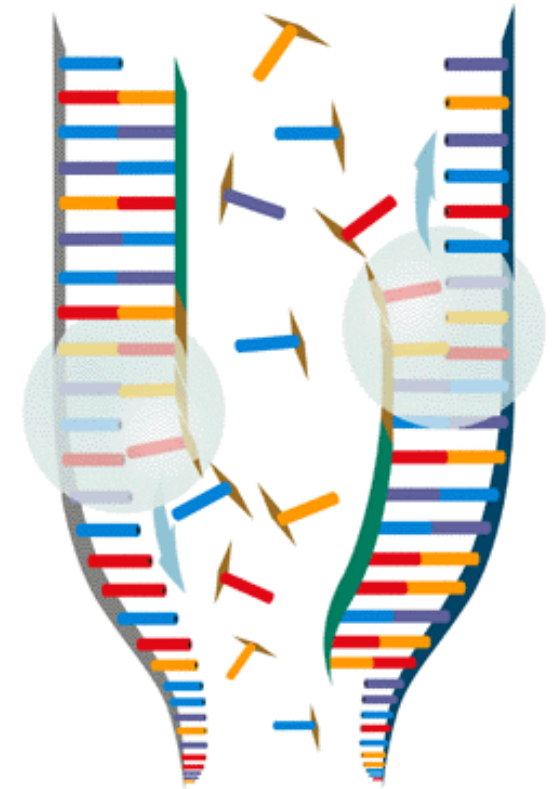


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# POLYMERASE CHAIN REACTION (PCR)

- Highly allergen-specific, highly sensitive
- Detects presence of DNA coding for allergen proteins
- Costly equipment, specialized, longer protocol
- Does not directly detect allergenic proteins
  - some processing results in DNA↓, protein↑



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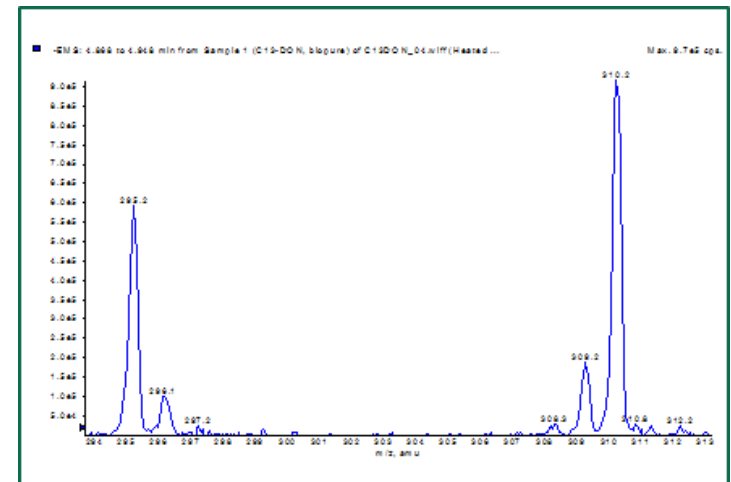
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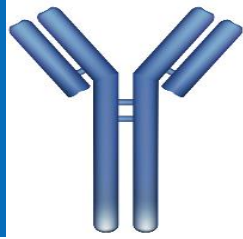
# LIQUID CHROMATOGRAPHY TANDEM MASS SPECTROMETRY (LC-MS/MS)

- Highly specific, extremely sensitive, quantitative
- Detects peptide fragments of allergenic proteins
- Equipment costly, large and highly specialized
- Limited number of labs performing LC-MS/MS testing for food allergens
- Ability to detect heavily processed allergen protein fragments



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# IMMUNOASSAYS

- Allergen-specific, very sensitive
- Detects using antibodies against specific allergen protein
- Quantitative (ELISA) or qualitative (LFD)
- Can be used for both cleaning verification and validation (ingredients, finished product also)

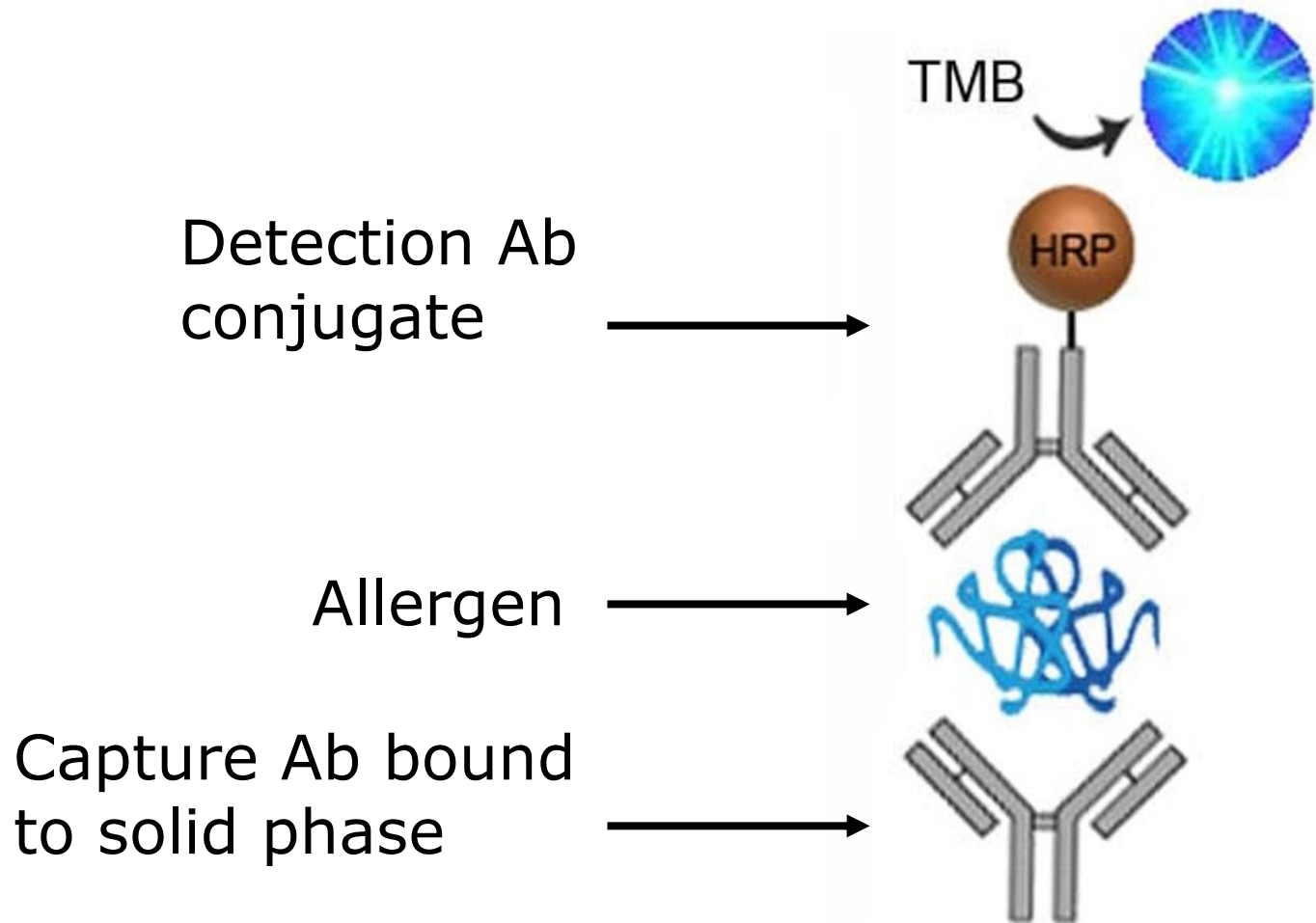


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# SANDWICH IMMUNOASSAY



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# ENZYME-LINKED IMMUNOSORBENT ASSAY (ELISA)

- Quantitative
- Some additional lab equipment, experience/training necessary
- 45-120 min test time
- Sample result extrapolated from standard curve
- Can test raw ingredient, finished product, swabs, rinses

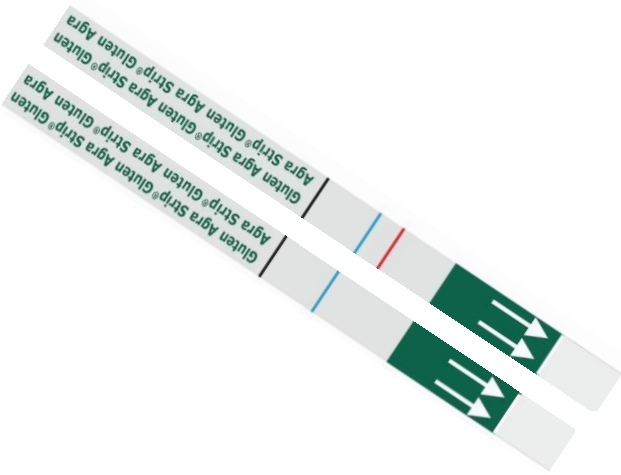


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# LATERAL FLOW DEVICE (LFD OR STRIP TEST)

- Qualitative:  $\geq$  or  $<$  LOD
- Rapid time to result
- No or little additional equipment
- Does not require lab

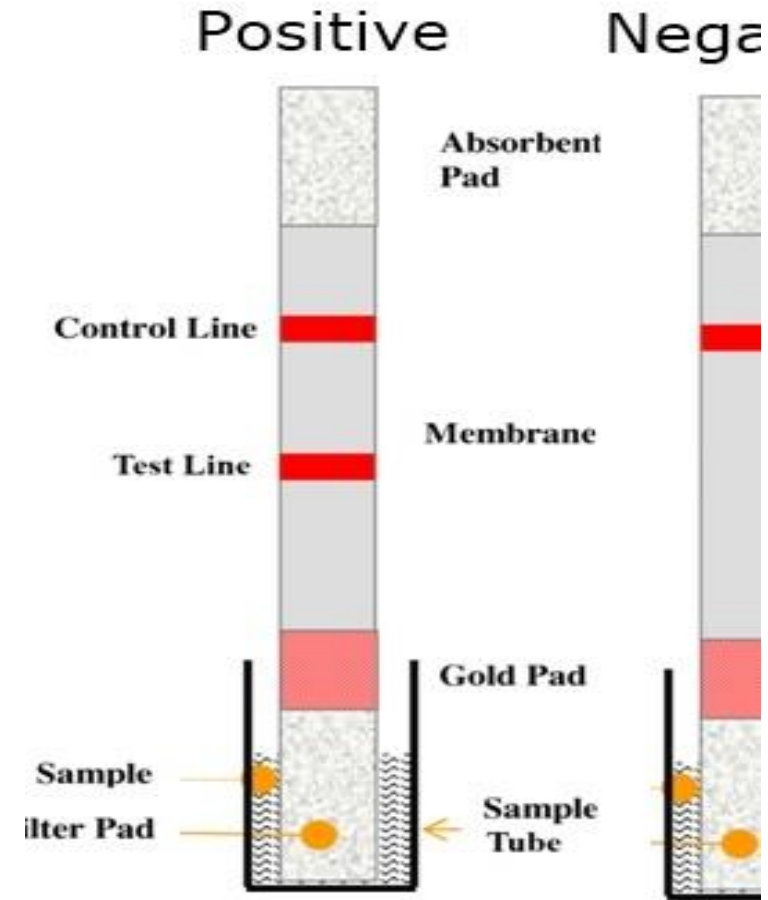


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# LATERAL FLOW DEVICE (LFD OR STRIP TEST)

- LFD Result Interpretation
  - Limit of detection (LOD) varies between allergens, manufacturers
  - Test line presence indicates allergen conc.  $\geq$  LOD
  - Test line absence indicates allergen conc.  $<$  LOD
  - Weak test line is still a positive



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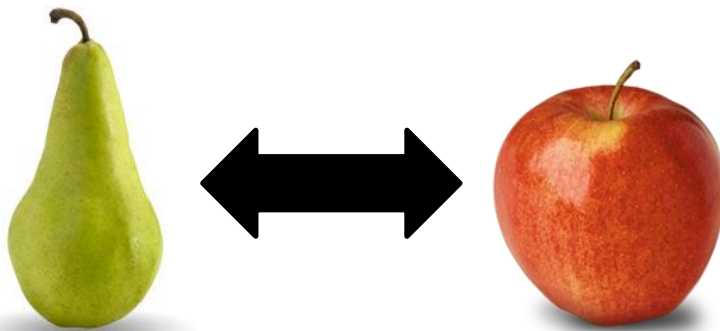


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# THE PROBLEM WITH ALLERGEN METHODS...

## Comparability!

- A variety of factors contributes to the different performance of test methods and test kits within the same method for the same allergen.



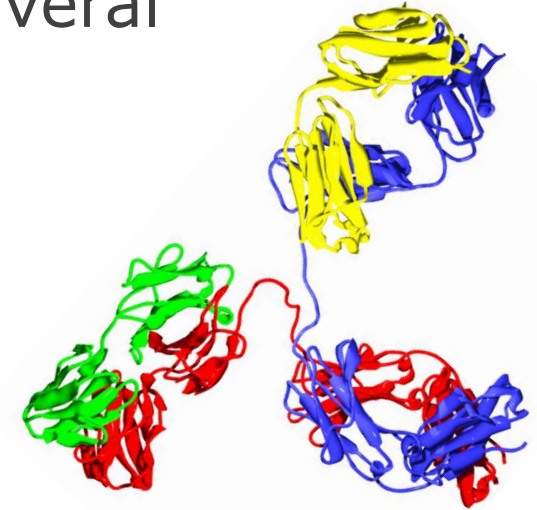
*Which kit's  
result is correct?*

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# FACTORS INFLUENCING IMMUNOASSAY MEASUREMENTS

- Antibodies
  - Monoclonal or polyclonal
  - Target of the antibody: single/several proteins, fractionated, modified, synthesized
  - Material used for immunization



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# TESTING FOR GLUTEN - ANTIBODIES

- mAb 401.21 (Skerritt)
  - 1<sup>st</sup> generation, raised against wheat
  - Majority of reactivity to glutelins,  $\omega$ -gliadin
  - Difficulty detecting barley hordein
- R5 (Mendez)
  - Raised against rye secalin, cross-reacts with wheat gliadin
  - Cross-reactivity to soy and lupin proteins reported
- G12 (Morón)
  - Raised against immunotoxic 33-mer of wheat gliadin
  - Cross-reacts strongly to homologous immunotoxic peptides in rye and barley
  - Cross-reactivity to oat avenin\*



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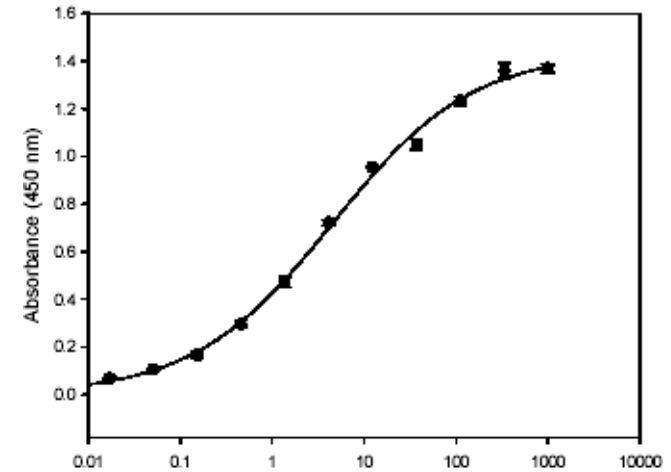
# FACTORS INFLUENCING IMMUNOASSAY MEASUREMENTS

- Calibrators

- Match the antibodies of the kit
- Not the same as sample material

- Extraction

- What is extracted (efficiency)?
- Antibody can only detect what has previously been detected



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# FACTORS INFLUENCING ELISA COMPARABILITY

- Limit of Detection (LOD)
  - Lowest amount of allergen to be distinguished from a true blank
  - LOD mostly determined in buffer
  - May be completely different in food samples
- Recovery
  - Ideally 80 – 120%
  - Not possible for all matrices (pH, salts, polyphenols,...)
  - Spiked vs. incurred samples



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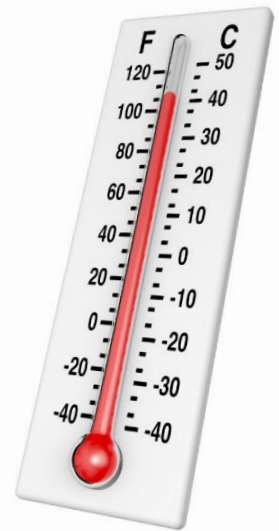
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# FACTORS INFLUENCING IMMUNOASSAY COMPARABILITY

- Immunological test system
  - Environmental influences
  - Experimental conditions
  - Stability affected by transport & storage conditions
  - Robustness, ruggedness



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# SAMPLING FOOD COMMODITY

- Sample from beginning, middle and end of lot
- Take a fairly large sample – at least 100 g, but 500 g to 1 kg is preferable
- Thoroughly mix sample to homogenize
- Subsample a smaller portion, and grind if necessary to as fine as possible
- Mix to homogenize subsample
- Take portions for testing according to manufacturer's assay instructions

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# IMMUNOASSAY MATRIX VALIDATION PARAMETERS

- Important to validate that matrix does not cross-react or interfere with assay (selectivity)
- Accuracy, precision and recovery across range of assay
- LOD/LOQ will vary, and should be established for each new matrix
- Incurred allergen reference material would be ideal, but very little available
- Often necessary to spike

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# SOURCES OF IMMUNOASSAY INTERFERENCE

- High protein concentration
- High lipid concentration
- High polysaccharide concentration
- Extremes of pH
  - May push past pH-buffering capacity of kit reagents
- Reactive biomolecules
  - Polyphenols
  - Tannins
  - Anthocyanins

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# MATRIX CHOICES: ALLERGEN MATERIAL

- Incurred
  - Better option – allergen processed along with food matrix
  - True evaluation of extraction efficiency
  - Difficult to design/predict allergen levels precisely
  - Challenge for method developer
  - Some incurred RM available for purchase
  - No CRM

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# MATRIX CHOICES: ALLERGEN MATERIAL

- Spike
  - Necessary for matrices which cannot be produced as incurred
  - Over-recovery of spike
  - Useful for measuring precision of detection portion, interference
  - Allergen extract vs whole allergen
  - Homogeneity



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# SUMMARY

- **Understand that every gluten detection method has strengths and weaknesses...no perfect method**
- **Know capabilities and limitations of any detection method prior to implementation**
- **Validate that method will detect gluten in your matrices**
- **When in doubt, contact tech services dept. of the kit manufacturer**

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# THANK YOU!



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